

# 10 Solutions and tips for bioimage figures

Helena Jambor, TU Dresden, Germany

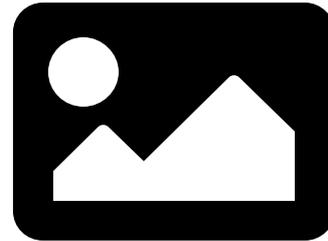
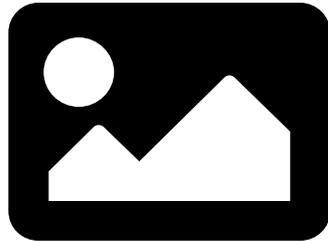
[helena.jambor@tu-dresden.de](mailto:helena.jambor@tu-dresden.de)



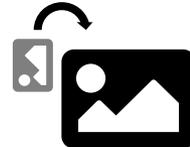
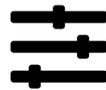
@helenajambor

1

# Make image copy



Work only on copy



Save original microscope image

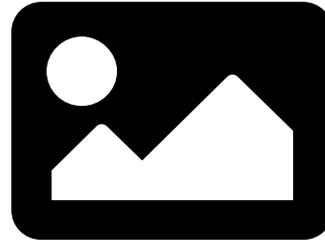


@helenajambor

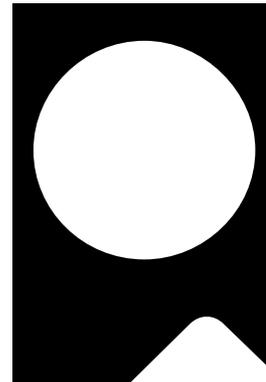
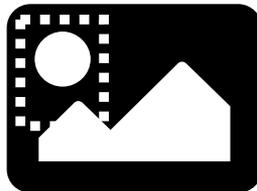
2

## Magnification

What size on final page?



Crop, show zoom, always indicate position!



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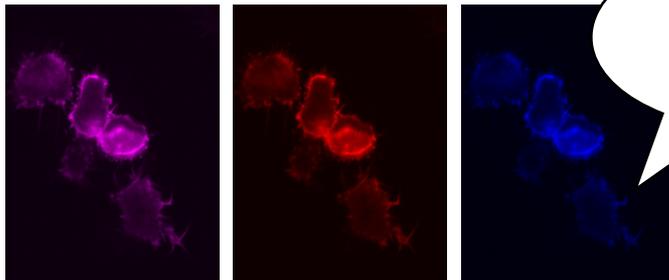
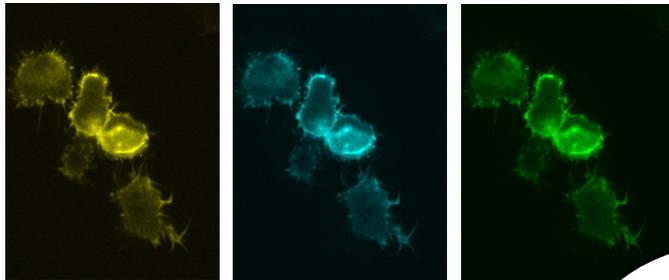
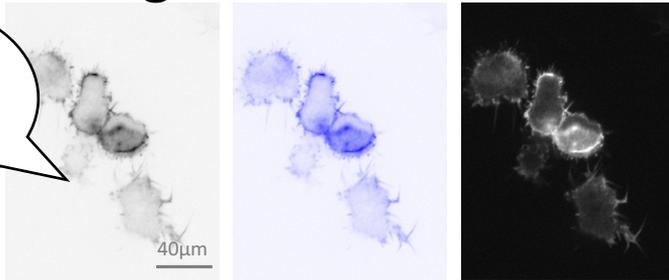
3

# Colors: 1-color image

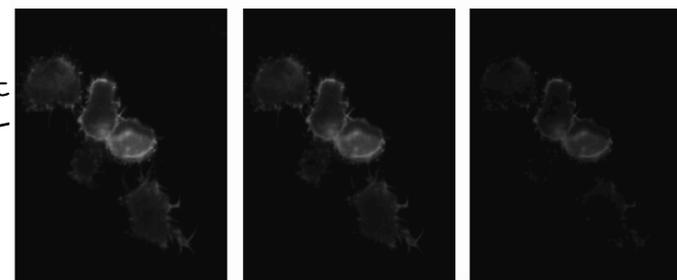
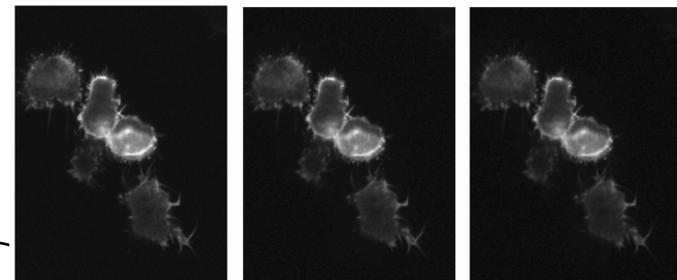
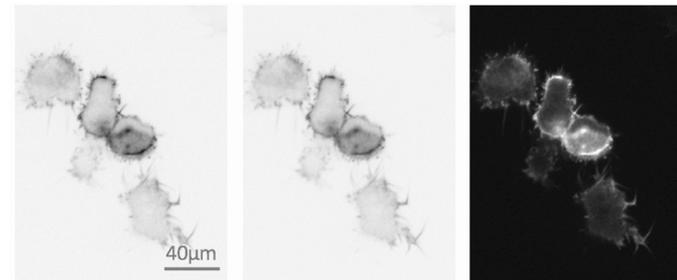
High contrast of signal on background color

Test visibility: greyscale

😊  
better



☹️  
poor contrast



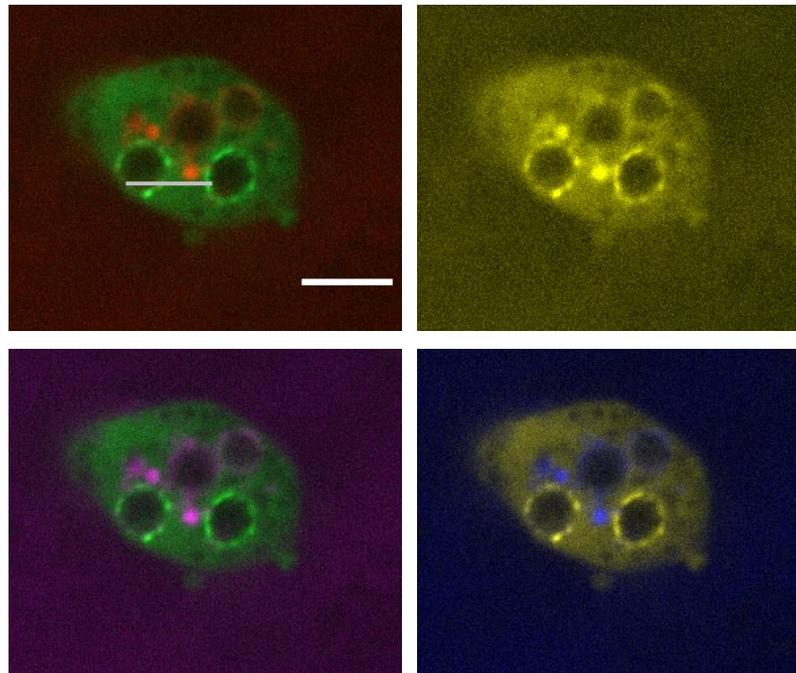
4

# Colors: 2-color image

## Be colorblind safe!

Normal vision

Deuteranopia



☹️  
not  
accessible

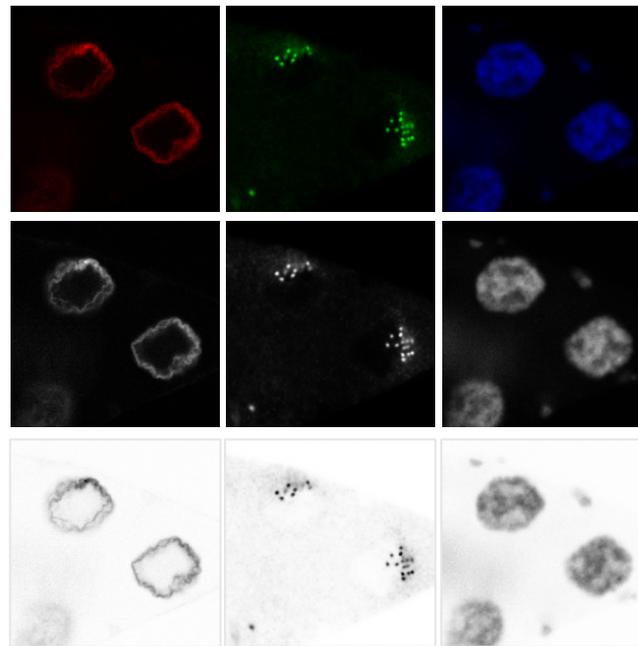
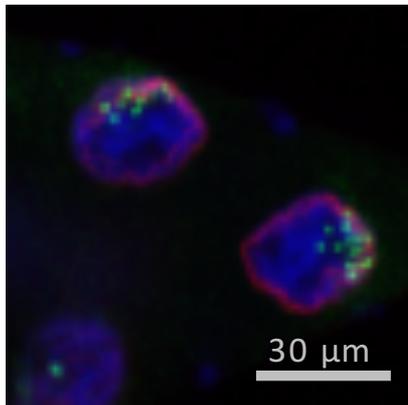
😊  
better

5

## Colors: 3-color image

Be colorblind safe!  
(You will struggle... )

Split channels, better in greyscale

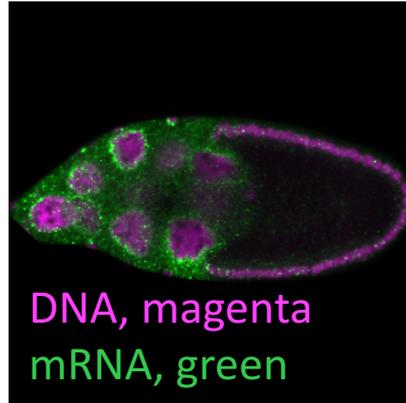
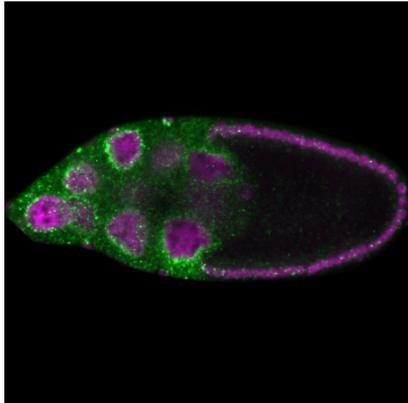


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# Annotate colors

Explain all colors used!



DNA, magenta  
mRNA, green

Annotation  
also color  
blind safe!  
😊

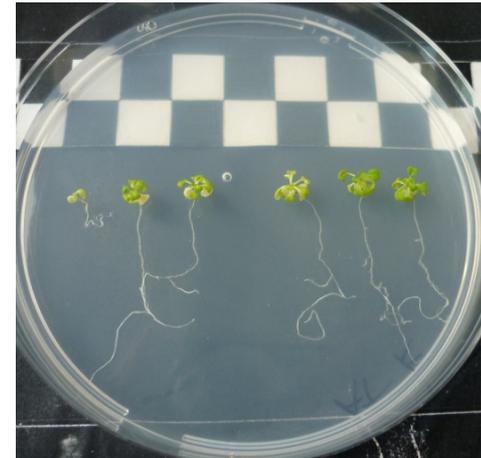
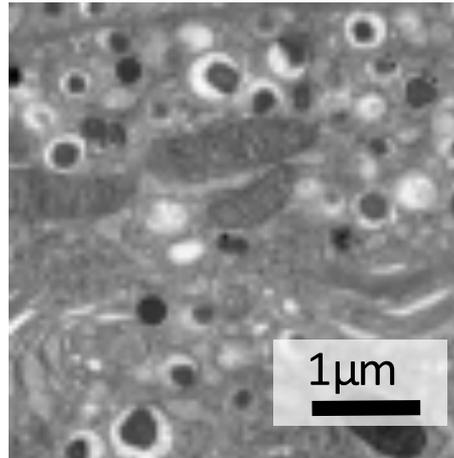
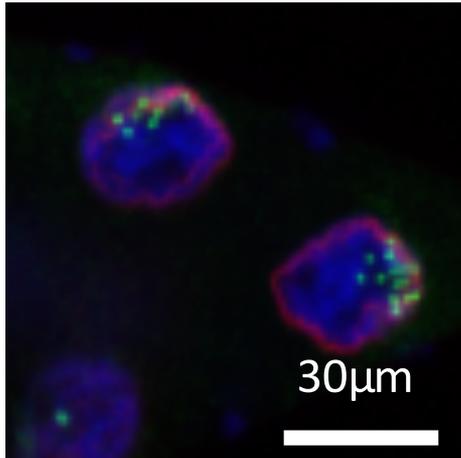


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## Annotate scale

Every scientific image need scale information



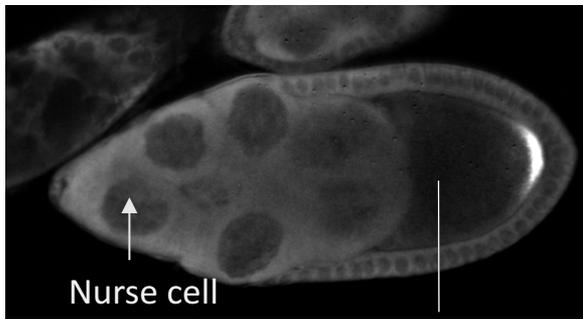
\*\*\* Magnification statement problematic



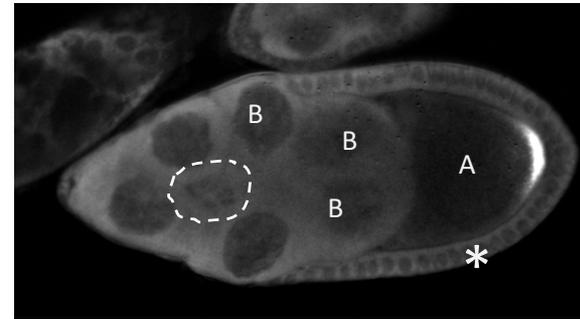
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# Annotate features

Lines, Arrows, Regions-of-interest, Symbols, Lettercode



Oocyte

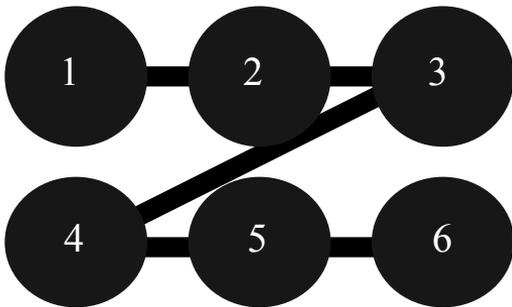


A Oocyte  
B Nurse cells  
\* Somatic Cells

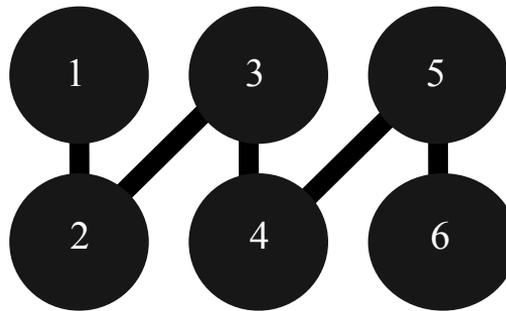


## Audience understands reading direction of images

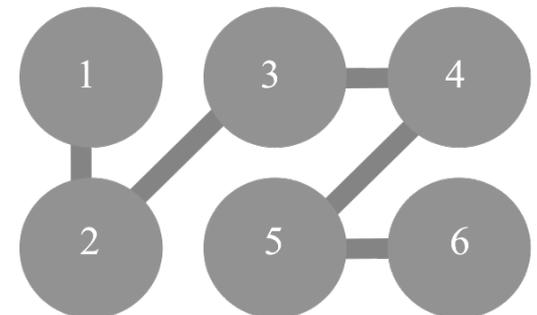
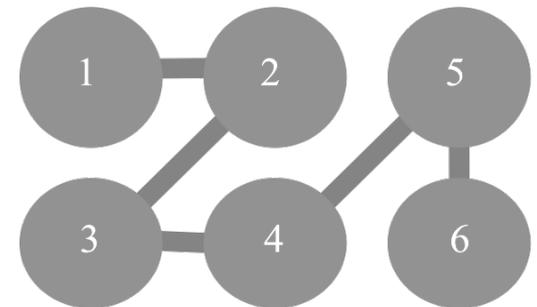
Images in rows



Images in columns



Don't mix!



No image is self-explanatory

Blog Total Internal Reflection by Brooke:

<https://totalinternalreflectionblog.com/2021/08/11/the-devil-in-the-details-a-short-guide-to-writing-figure-legends/>



# Help is near *Cheat sheets for novice users*

Christopher Schmied & Helena Jambor, F1000R, 2020

## Processing images for papers & posters: CHEAT SHEET

Download: [www.fiji.sc](http://www.fiji.sc)

Update and install:

**Help > Update...**

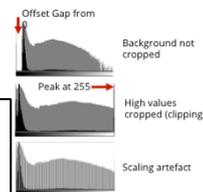
**ImageJ Updater > Manage update sites then add specific site**

**Open & Save**

**File > Open...** or **Ctrl + O** or **Drag & Drop**



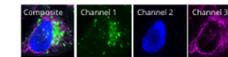
Typical problems in images:



### Color

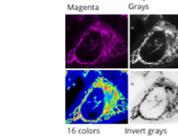
**TIPP** In composite images, switch between channels for viewing details:  
**Image > Color > Channels Tool**

To adjust color, split composite image to separate channels:  
**Image > Color > Split**



Change LUTs to preset color/colorscheme:  
**Image > Lookup Tables...**  
(pick color e.g. *grays, magenta, 16 colors*)

Invert for better visibility:  
**Edit > Invert...** - **Ctrl + Shift + I**



### Annotate

Set scale:

**Analyze > Set Scale...**

**Analyze > Tools > Scale Bar...**

Width: 1/5/10 steps

Color: highest contrast to image, if necessary add background

Overlay: separate layer from image (lost in png)

Hide text and add later if resolution of image too low



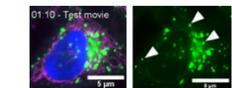
**TIPP** overlay may be turned on/off:

**Image > Overlay > Hide/Show Overlay**

Add text and time labels:

**Image > Stacks > Label...**

**Image > Annotate > Arrow...**  
arrows can be moved/rotated/shortened



**TIPP** manage multiple ROIs and Labels:

**Analyze > Tools > ROI Manager**

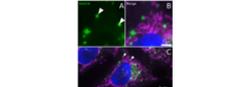
### Layout

Plugins to create multi-image panels with annotations for publications:

Figure: <https://image.net/FigureJ>

ScientificFig:

<https://jgr.great-clermont.fr/abimouse/software/>



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hambor@gmail.com, Schmied@mp-berlin.de

## Publishing images for papers & posters: CHEAT SHEET

### Image check list

Before publishing, ensure images are informative, truthful, and legible.

**Colors visible?** Grayscale and dark colors on light background have highest contrast. Test visibility in color blind mode e.g. with [colororacle.org](http://colororacle.org)

**Annotations complete?** Explain annotations and abbreviations in figure legend. Hint: best avoid abbreviations.

**Image and text size on page:** Test print figures: are image features visible is text legible?

**Image resolution:** Be careful with image compression when saving, avoid pixelated images.

**No manipulation!** No individual image feature may be enhanced, obscured, removed, or introduced.



### Magnification

Evidences in images must be visible to be convincing. Scientists should decide on the necessary detail and fill the frame to maximize the information/pixel.

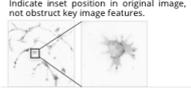


Tissue information Cellular information Subcellular information



### Zoom, Insets

Insets or zoomed regions-of-interest are used when two magnifications are needed. Indicate inset position in original image, do not obstruct key image features.

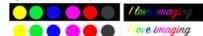


### Color

Color or not?

- Photo of natural appearance, color helps.
- Micrograph of a colored stain (e.g. histology), color helps.
- Micrographs of fluorescent stain, imaged in grayscale, consider grayscale.
- Micrographs of 2-3 fluorescent stain color, use color-blind safe combination.
- Electron micrographs, grayscale by default, use grayscale.

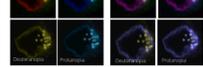
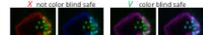
Which color? Visibility depends on color lightness and background color.



To a color blind, some colors are not distinguishable, often red and green (right).



Pick color blind safe combination:



Explain colors

• red color-blind safe, no labels  
• Channel shown separately  
• Color-blind safe scheme



**Resources for color** Channel colors are adjustable in ImageJ/Fiji, referred to as lookup tables (LUT). Colors can also be inverted and new LUTs defined.

**TIPP** Highest contrast: dark objects on light background and in grayscale.

### Annotate

**Scale information** Every image needs a scale to relate it to reality. Options:

- Scale bar of known length, if possible dimensions annotated in image.
- Magnification of set-up indicated, less intuitive and error-prone.
- Tape measure/ruler included next to object in photo.
- Every day item shown next to object.

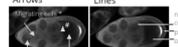


**Annotation on a busy background**



**TIPP** in Fiji to insert a very thin scale, then add rectangle as scale bar with figure design software to better adjust thickness/point size.

**Point out details**



**Region of interest** Letter code



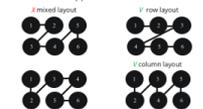
**Use** Arrows: Point to structure, show directional movement (!)  
ROI: Point to entire structure  
Lines: Label at line end  
Letter: Label many features

**Tips** Align, avoid arrow overlap  
Choose suitable point size for dashed lines  
Align, avoid line crossings  
Choose suitable font, label, size

**TIPP** Avoid overlapping annotations and obstruction of key image features. Annotation colors should be color blind safe and aligned.

### Layout

**Reading direction** For multi-panel figures, choose either column or row layout for best readability. Mixed layouts are hard for audience and appear cluttered.



**Place panels on grid** with white space and imaginary lines to orient audiences. Panels should not extend beyond grid lines.



**TIPP** sketch, draw, use post-it & pen until it works. Only then start an electronic version.

### Figure legend

Figure legends inform briefly about result, experiment, and figure elements.

- Include header, check journal guidelines
- Annotate images directly, if not possible indicate scale and colors in legend.
- State species (e.g. with BBID), tissue/cell type and treatment (e.g. antibody).
- Avoid method details and discussion.

**TIPP** Get feedback! Test your figure by asking a colleague to explain it back to you: this quickly uncovers missing text, insufficient annotations etc.

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hambor@gmail.com, Schmied@mp-berlin.de  
ImageJ: Helena Jambor, Arndt Müller (BM)  
Scopus ID: Helena Jambor, Template: BioRxiv



@helenajambor

# Publishing images for papers & posters: CHEAT SHEET

## Image check list

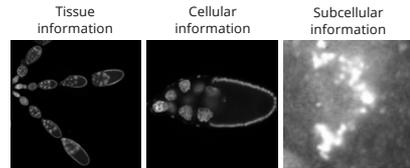
Before publishing, ensure images are informative, truthful, and legible.

- ❑ **Colors visible?** Grayscale and dark colors on light background have highest contrast. Test visibility in color blind mode e.g. with [colororacle.org](http://colororacle.org)
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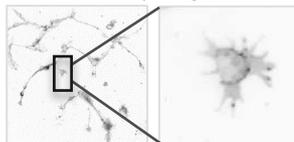
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- Photo of natural appearance, color helps.
- Micrograph of a colored stain (e.g. histology), color helps.
- Micrographs of fluorescent stain, imaged in grayscale, consider grayscale.
- Micrographs of 2-3 fluorescent stain color, use color-blind safe combination.
- Electron micrographs, grayscale by default, use grayscale.

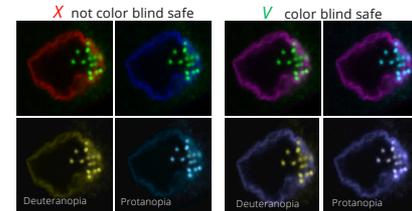
**Which color?** Visibility depends on color lightness and background color.



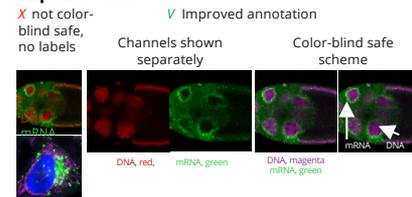
To a color blind, some colors are not distinguishable, often red and green (right).



Pick color blind safe combination:

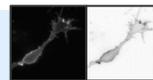


### Explain colors



**Resources for color** Channel colors are adjustable in ImageJ/FIJ, referred to as lookup tables (LUT). Colors can also be inverted and new LUTs defined.

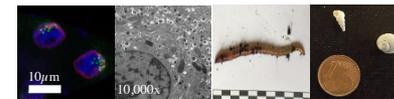
**TIPP** Highest contrast: dark objects on light background, and in grayscale.



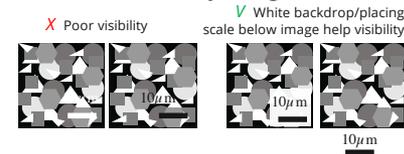
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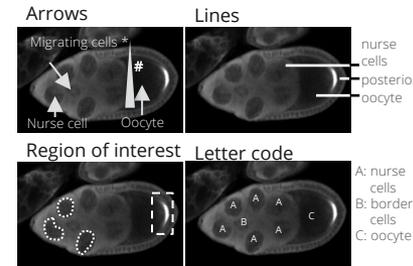


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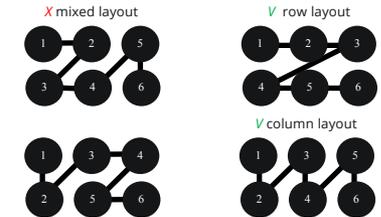


Use	Tips
<b>Arrows</b>	Point to structure, show direction/movement (*) Align, avoid arrow crossings
<b>ROI</b>	Point to entire structure Choose suitable point size for dashed lines
<b>Lines</b>	Label at line end Align, avoid line crossings
<b>Letter</b>	Label many features Choose suitable font (sans serif)

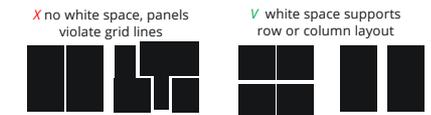
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- Annotate images directly, if not possible indicate scale and colors in legend.
- State species (e.g. with [RRID](#)), tissue/cell type and treatment (e.g. antibody).
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Download: [www.fiji.sc](http://www.fiji.sc)

Update and install:

**Help > Update...**

**ImageJ Updater > Manage update sites**  
then **add specific site**



## Open & Save

**File > Open ...** or **Ctrl + O** or **Drag & Drop**

Special Formats:

**Plugins > Bio-Formats > Bio-Formats Importer**

Review existing metadata:

**Image > Properties...** - **Ctrl + Shift + P**

**File > Save As ...**

**TIFF:** analysis, quantifications

**PNG:** presentations, figures

**AVI:** movies, animations, GIFs

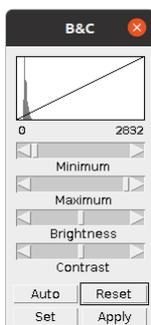
**TIPP** duplicate image before processing:

**Image > Duplicate...** - **Ctrl + Shift + D**

## Brightness & Contrast

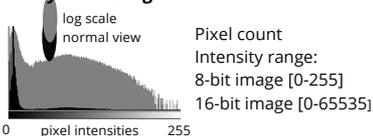
**Image > Adjust > Brightness/Contrast...**  
**Ctrl + Shift + C**

- Auto: saturates the image by 0.35%
- Reset: to min & max or 0-255 for 8-bit
- Set: fixed values - use for comparisons
- Apply: histogram stretch using set min & max. **Use with caution!**



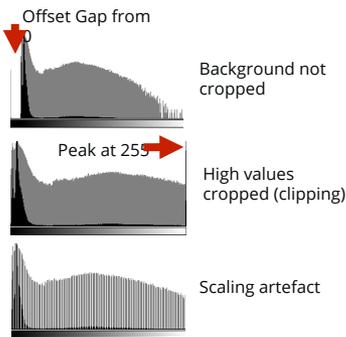
Check for problems with intensity sampling:

**Analyze > Histogram...** - **Ctrl + H**



**TIPP** during acquisition use ~75% of the intensity range histogram to avoid saturation or too low signal capture.

Typical problems in images:



## Image Processing

Gaussian blur:

**Process > Filters > Gaussian Blur...**

Projection:

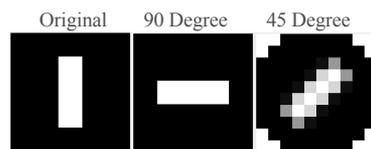
**Image > Stacks > Z Project...**

## Rotation & Resizing

**Image > Transform > Rotate 90 Degrees...**

**Image > Transform > Flip...**

Ideally measure before rotation



Save images in smaller format and size:

**Plugins > Example > Downsample...**

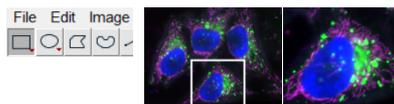
Avoid upsampling

## Cropping

Draw selection in toolbar

**Image > Crop...** - **Ctrl + Shift + X**

**Image > Duplicate...** - **Ctrl + Shift + D**



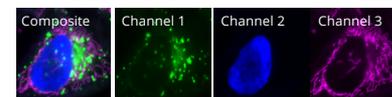
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**Image > Color > Channels Tool**

To adjust color, split composite image to separate channels:

**Image > Color > Split**



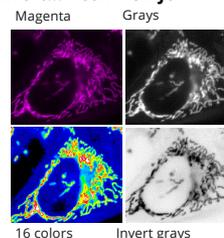
Change LUTs to preset color/colorscheme:

**Image > Lookup Tables...**

[pick color e.g grays, magenta, 16 colors]

Invert for better visibility:

**Edit > Invert...** - **Ctrl + Shift + I**



Merge channels with predefined LUT:

**Image > Color > Merge Channels...**

tick 'ignore source LUTs' to merge with custom LUTs

tick 'make composite' to retain bit-depth  
Create your own LUTs:

**Image > Color > Edit LUT...**

**TIPP** test color blind safety: Most common form color blindness: deuteranopia. Test general visibility in grayscale or "Monochromacy".

Required: RGB image.

**Image > Color > Stack to RGB...**

**Image > Color > Simulate Color Blindness...**

## Annotate

Set scale:

**Analyze > Set Scale...**

**Analyze > Tools > Scale Bar...**

- Width: 1/5/10 steps
- Color: highest contrast to image, if necessary add background

Overlay: separate layer from image (lost in png)

Hide text and add later if resolution of image too low

**TIPP** overlay may be turned on/off:

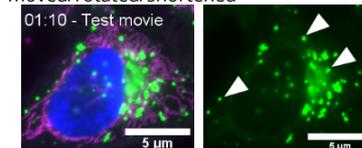
**Image > Overlay > Hide/Show Overlay**

Add text and time labels:

**Image > Stacks > Label...**

**Image > Annotate > Arrow...**

arrows can be moved/rotated/shortened



**TIPP** manage multiple ROIs and Labels:  
**Analyze > Tools > ROI Manager**

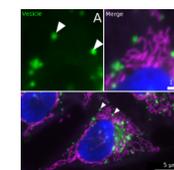
## Layout

Plugins to create multi-image panels with annotations for publications:

FigureJ: <https://imagej.net/FigureJ>

ScientiFig:

<https://grr.gred-clermont.fr/labmirouse/software/>

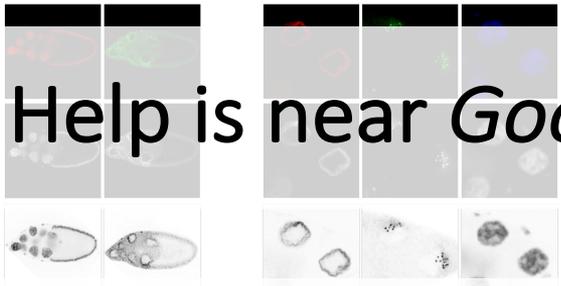


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hjambor@gmail.com, Schmieid@fmp-berlin.de

2. Split channels: colorblind safe, some colors have low contrast (see Fig. 6)

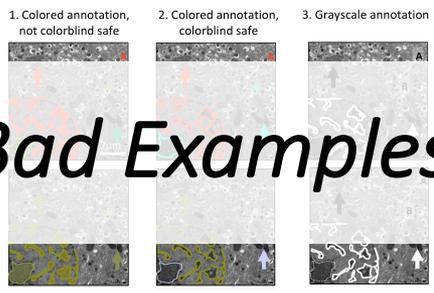
3. Show split channels in grayscale: colorblind safe, high contrast

4. Show split channels in inverted grayscale: colorblind safe, highest contrast

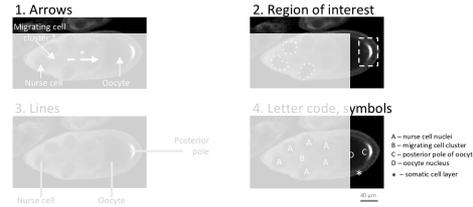


# Help is near *Good/Bad Examples*

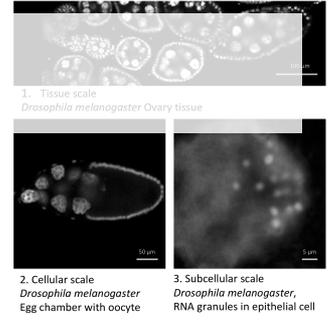
Example



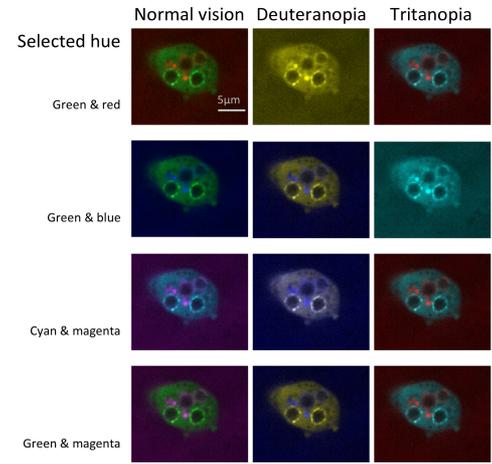
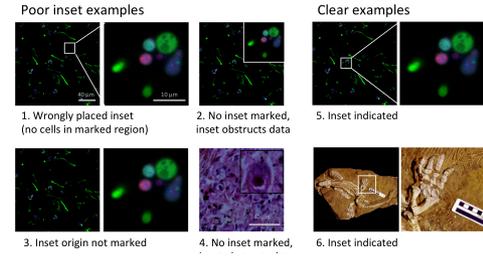
## Annotation strategies



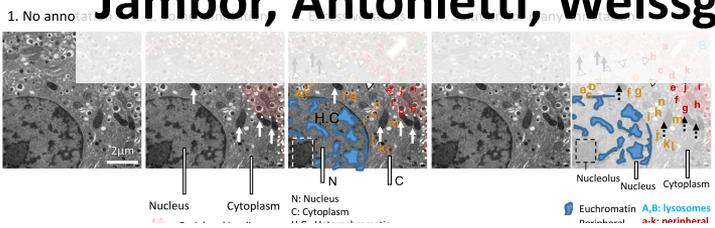
A Magnification/zoom must match message



B Insets allow readers to see more than one scale

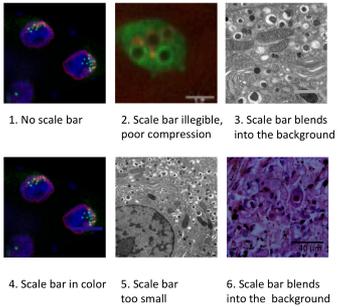


## Jambor, Antonietti, Weissgerber et al PLOS Bio 2021

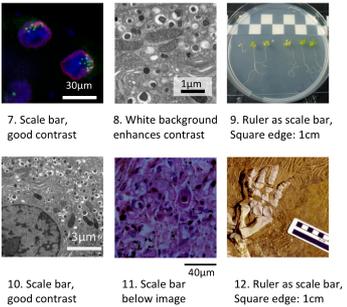


Legend: N: Nucleus, C: Cytoplasm, H.C: Heterochromatin, A, B: lysosomes, a-k: peritubular

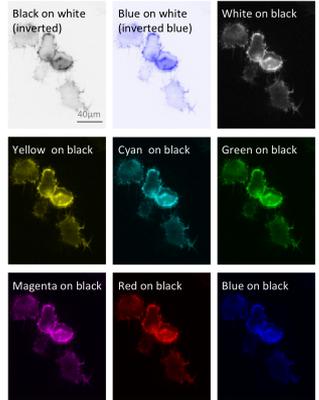
Poor scale bar examples



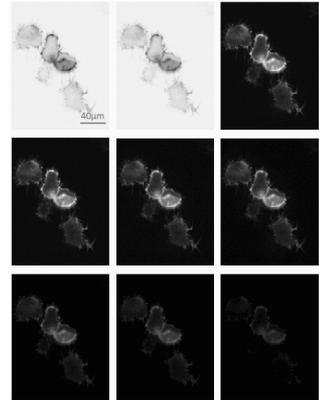
Clear scale bar examples



Color images



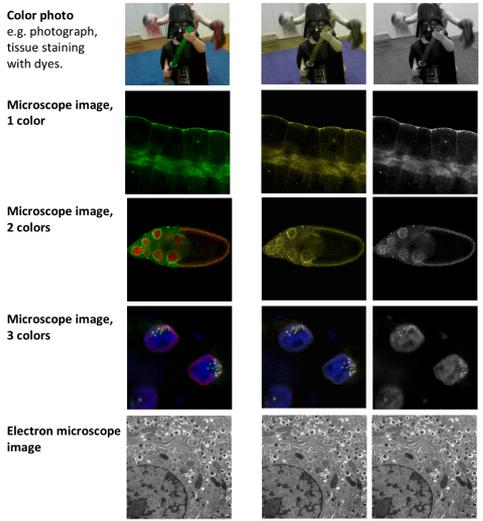
Grayscale test for visibility



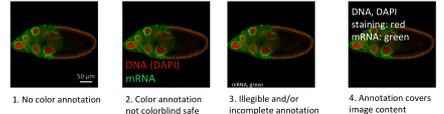
Situation

Example

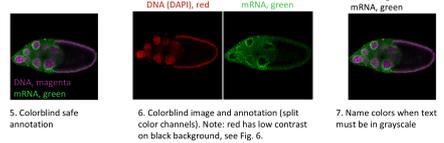
Visibility tests



Poor color annotation

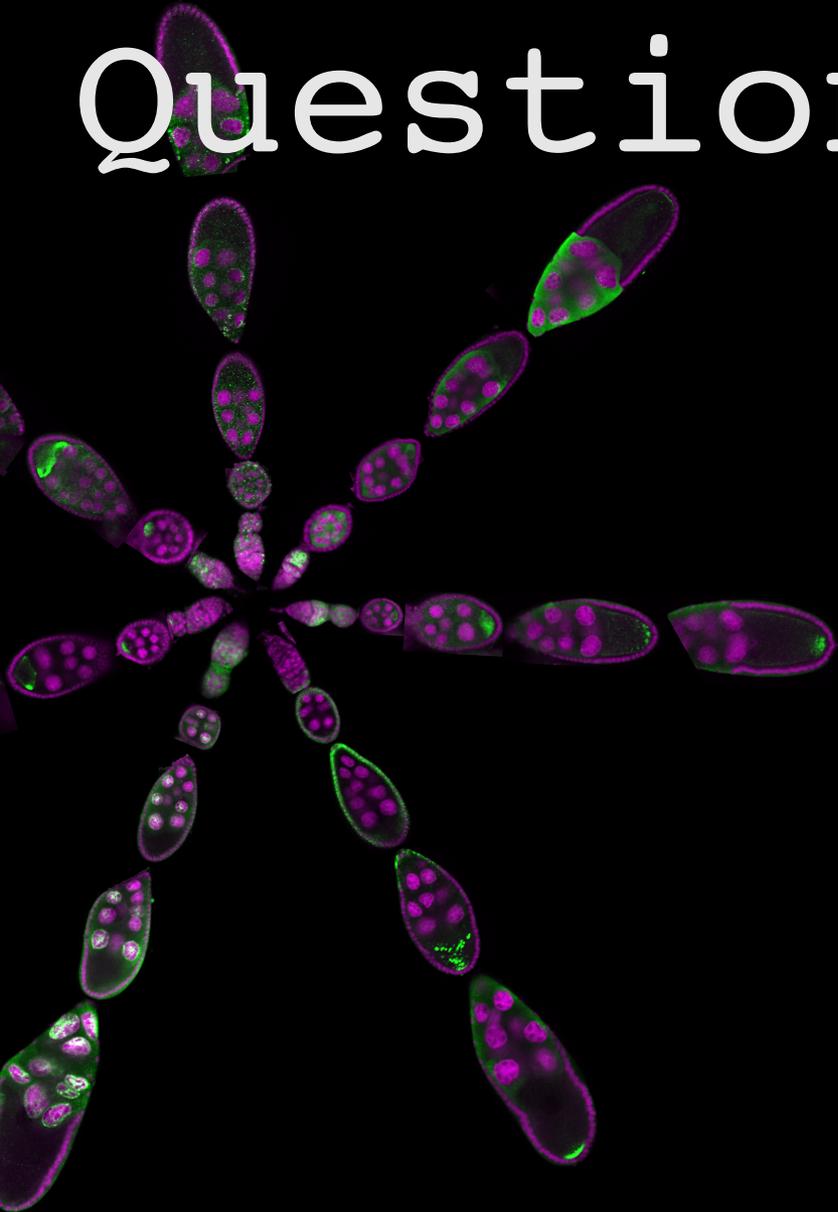


Clear color annotation



# Question

# time!



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